Ring Opening Reactions of 6-Deoxy-9-deoxo-9a-aza-9a-homoerythromycin A 6,9-Cyclic Imino Ether

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(Received for publication April 15, 1996)

Treatment of 6-deoxy-9-deoxo-9a-aza-9a-homoerythromycin A 6,9-cyclic imino ether with hydroxylamine hydrochloride led to the cleavage of the C9/9a-N imino bond with formation of C-9 oxime. Base catalyzed internal acylation of the newly introduced C-10 *primary* amino group by C-1 ester followed by catalytic hydrogenation and reductive methylation afforded a series of novel C-1 amides, the first representatives of linear 9a-azalides in which the C-10 to C-15 western fragment is inverse bound to the C-1 carbon atom.

6-Deoxy-9-deoxo-9a-aza-9a-homoerythromycin A 6,9-cyclic imino ether (3) is an important intermediate in the synthesis of azithromycin (1), the first example of a new class of azalide antibiotics differing structurally from erythromycin A by an 9a-aza substituted, ring expanded, 15-membered macrocyclic framework^{1,2)}. The crucial step in the synthesis of 1 is the stereospecific Beckmann rearrangement of erythromycin A 9(E)-oxime³⁾. Similar experiments with the 9(Z)-isomer⁴⁾ led to a series of novel 8a-aza-8a-homoerythromycin analogs. Among them, 8a-methyl derivative (2) was shown to exhibit in vitro activity equal to that of 1.

Recently, two different types of base catalyzed ring opening reactions of erythromycin A have been reported which led to formation of the C-1 carboxylate⁵⁾. Thus, in an aprotic solvent with strong base a retroaldol fragmentation gave a C-1 to C-10 segment which was

demonstrated to play an important role in the synthesis of two linear nitrogen containing fragments⁶). Such compounds represent a new approach to the synthesis of *chimeric* 9a- and 8a-azalides which have high structural homology to potent prototypes 1 and 2 in the critical eastern half of the molecule, but which have widely varying western portions derived from an essentially unlimited number of exogenous sources⁷). This communication describes the synthesis of a new 6,9-cyclic seco oxime starting from 6,9-cyclic imino ether 3 and its conversion into novel type of linear 9a-azalide structures (Scheme 1).

A well-known strategy for the synthesis of lactam oximes is the reaction of lactim ethers with hydroxylamine⁸⁾. This reaction became important when the lactam oximes were found to undergo the Beckmann rearrangement yielding ureas. In the course of our efforts to prepare the lactam oxime from 6,9-cyclic imino ether 3, we have discovered a simultaneous cleavage of C-9/9a-N double bond which leads the formation of the seco oxime (4a). This reaction step was accomplished in MeOH by condensation of 3 with hydroxylamine hydrochloride in the presence of Na₂CO₃ (91% yield).

FAB-MS showed the molecular ion peak at m/z 764 (MH⁺) satisfying the proposed molecular formula $C_{37}H_{69}N_3O_{13}$. The typical absorption at 1580 cm⁻¹ in the IR spectrum and the positive color obtained from the reaction with ninhydrin suggested the presence of the amino functional group. The shieldings observed for both the 10-H proton (-0.69 ppm) and C-10 carbon signals (-6.0 ppm) in the ¹H and ¹³C NMR spectra of **4a** with respect to 399 indicated that this group may be attached to the C-10 carbon. The hydroxyimino proton at C-9 $(\delta_{\rm H} = 9.33, \ {\rm D}_2{\rm O} \ {\rm exchangeable})$ was deduced from ¹H NMR in DMSO- d_6 and significant change of 13 C chemical shifts for C-9 (-2.6 ppm) and C-8 (-3.2 ppm). Further examination of the ¹H NMR data showed that apart from a change of some coupling constants (${}^3J_{2,3}$ $2.4 \rightarrow 1.9$, ${}^{3}J_{4,5}$ $7.3 \rightarrow 8.3$, ${}^{3}J_{10,11}$ $10.1 \rightarrow 2.7$ Hz) and the deshielded 3-H proton resonance (+0.68 ppm) which indicated a different conformation, the shift values for both 3 and 4a are very similar. These results implied

Chart 1

Scheme 1. Synthetic pathways of $4a \sim 7$.

Reagents: i) NH₂OH, Na₂CO₃, MeOH; ii) Ac₂O, Py; iii) basic solution; iv) PtO₂, H₂, AcOH; v) HCOH, HCOOH, CHCl₃.

the rupture of the 15-membered macrocyclic ring between C-10 and 9a-N centers giving an unexpected seco derivative with the oxyimino function at C-9 and the terminal amino group at C-10. Consequently, the acetylation of 4a (Ac₂O, Py, 7 days, RT) afforded the expected pentaacetate (4b) which in the ¹H NMR spectrum revealed five acetyl signals at $\delta_{\rm H}$ =2.16, 2.13, 2.12, 2.05 and 1.96. The considerable deshielding of the 10-H signal ($\delta_{\rm H}$ = 3.05 \rightarrow 4.48), which was correlated with the resonance at $\delta_{\rm H}$ = 6.15 in the ¹H-¹H COSY experiment, confirmed the presence of a new amido group at C-10. The deshieldings of 2'-H ($\delta_H = 3.21 \rightarrow 4.81$), 4"-H $(\delta_{\rm H} = 3.04 \rightarrow 4.69)$, 11-H proton $(\delta_{\rm H} = 3.49 \rightarrow 4.67)$ and C-9 $(\delta_C = 163.7 \rightarrow 167.4)$ carbon resonances compared to those of 4a, clearly show that 4b is 2',4",11-O,10-N-tetraacetyl-9-deoxo-6-deoxy-6,9-epoxy-8(R)-methyl-10-amino-9,10secoerythromycin A 9(E)-acetoxime.

During the purification by column chromatography on silica gel with CHCl₃-MeOH-NH₄OH, 6:1:0.1 as the solvent, 4a was found to be converted quantitatively to (5a). The same compound was also formed when 4a was kept under basic conditions at room temperature.

The structural assignment of 5a was based on the following chemical and spectral evidence. FAB-MS showed the molecular ion peak at m/z 764 (MH⁺) which was identical with that of 4a. The IR spectrum showed new amide absorptions at 1650 and 1530 cm⁻¹ and loss of ester and amine bands at 1720 and 1580 cm⁻¹, respectively. Further, the new signal at $\delta_{\rm H}$ =7.53 in the ¹H NMR spectrum attributed to an amide proton, gave a cross peak with 10-H signal in the 2D homonuclear COSY spectrum[†]. As compared to **4a**, the deshielding of 10-H (+1.06 ppm), typical for acylation of an amino group, together with concomitant shielding of 13-H $(-1.56 \,\mathrm{ppm})$ and the more deshielded C-13 carbon $(+4.9 \,\mathrm{ppm})$, led us to the hypothesis of a base-induced intramolecular transacylation between C-10 primary amino group and C-1 ester causing the inversion of a C-10 to C-15 western portion of the molecule. Therefore, the new amide group is attached to the C-1 carbon in 5a. This explanation is supported by a long-range correlation between C-1 and proton signal 13-H in the COLOC spectrum of 4a, a connectivity which was absent in the corresponding spectrum of 5a. Additional evidence

[†] The numbering system of the C-10 to C-15 fragment after the inversion in seco amides $5a \sim 7$ is the same as that in 3.

Table 1. Characteristic ¹H (300 MHz, in CDCl₃) and ¹³C (75 MHz, in CDCl₃) NMR data for the seco compounds 4a ~7.

Position ^a -	δ_{H} (ppm)					$\delta_{ m C}$ (ppm)				
	3	4a	5a	6	7	3	4a	5a	6	7
1						178.1	175.5	174.4	174.1	174.7
2	2.73	2.84	2.53	2.52	2.57	44.3	43.4	42.8	41.6	41.7
3	3.92	4.60	4.20	4.26	4.27	76.3	76.7	79.8	79.7	79.8
4	1.88	1.82	1.97	2.01	2.16	42.8	39.7	38.6	37.5	37.3
5	3.93	3.90	3.66	3.41	3.41	79.0	81.9	86.3	92.3	90.5
6						87.7	88.5	90.3	74.6	73.7
7a	2.01	2.08	2.10	2.20	1.50					
7b	1.57	1.88	1.79	1.34	1.15	36.9	39.5	41.0	42.8	44.2
8	2.80	2.92	3.04	1.85	2.01	35.1	31.9	32.9	31.0	26.4
9a				2.62	2.52					
9b				ND	2.20	163.7	161.1	162.0	49.1	68.2
10	3.74	3.05	4.11	4.17	4.18	52.7	46.7	48.6	49.2	48.6
11	3.67	3.49	3.79	3.76	3.77	72.0	72.5	75.I	74.8	74.6
12						75.1	73.5	74.9	75.1	74.8
13	4.94	4.78	3.22	3.17	3.18	77.6	78.1	83.0	83.8	83.3
14a	1.89	1.87	1.59	1.55	1.56					
14b	1.48	1.51	1.33	1.34	1.37	21.5	21.3	24.8	25.0	24.9
15	0.89	0.87	1.04	1.05	1.04	11.1	10.6	11.5	11.5	11.5
16	1.19	1.28	1.12	1.10	1.09	13.1	11.0	9.9	8.6	11.4
17	1.12	1.17	1.10	1.07	1.09	9.2	10.7	10.9	10.7	8.2
18	1.43	1.58	1.52	1.30	1.31	25.8	24.8	23.9	23.6	25.2
19	1.21	1.22	1.23	1.01	0.98	18.3	18.1	15.8	20.6	21.3
20	1.26	1.18	1.25	1.25	1.24	16.6	15.8	15.6	16.1	15.7
21	1.09	1.06	1.14	1.14	1.13	17.6	17.1	21.1	21.6	21.3
9aNMe					2.27					45.3
CONH			7.53	7.52	7.26					

^a The numbering system of the C-10 \sim C-15 fragment after the inversion in seco amides $5a \sim 7$ is the same as that in 3.

for the proposed structure of 6,9-cyclic seco oxime **5a** was obtained by acetylation giving (**5b**). The location of five acetyl groups ($\delta_{\rm H}\!=\!2.15,\ 2.12,\ 2.12,\ 2.06$ and 2.01) was pointed out by the deshieldings of 2'-H ($\delta_{\rm H}\!=\!3.37\!\rightarrow\!4.80$), 4"-H ($\delta_{\rm H}\!=\!2.96\!\rightarrow\!4.69$), 11-H ($\delta_{\rm H}\!=\!3.79\!\rightarrow\!4.55$), 13-H $\delta_{\rm H}\!=\!(3.22\!\rightarrow\!4.94)$ proton and C-9 ($\delta_{\rm C}\!=\!162.0\!\rightarrow\!167.2$) carbon signals.

6,9-Cyclic seco oxime 5a was unreactive towards sodium borohydride in methanol or high pressure hydrogenation using Pt/C in 96% ethanol. However, the catalytic reduction of 5a using PtO₂ in acetic acid under 7×10^6 Pa H₂ for 10 hours produced the 9a-aza linear azalide (6) in 73% yield. The amine 6 was methylated under Eschweiler-Clark conditions to give the 9a-dimethyl derivative (7) (61% yield). The structures of 6 and 7 were determined by 2D NMR including COSY, HMQC and HMQC-TOCSY spectra and ultimately confirmed by FAB-MS giving the expected molecular ions at m/z 752 and 780, respectively. The 1 H and 13 C NMR chemical shifts for seco azalides $4a \sim 7$ are summarized in Table 1.

In summary, the ring opening reaction of 6,9-cyclic imino ether 3 with hydroxylamine is an efficient method for the synthesis of 9a-aza seco azalides with inversion of C-10 to C-15 western fragment, potential intermediates for new azalide antibiotics.

Acknowledgments

This work was supported in part by a grant from The Ministry of Science, Technology and Informatics, Republic of Croatia, project number 1-07-035 and the Fonds der Chemischen Industrie. We wish to thank Mrs. J. IVETIĆ for technical assistance.

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